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## Clinical Study

# Urinary Eosinophil Protein X in Children with Atopic Asthma

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The aim of this study was to investigate the relationship between urinary eosinophil protein X (uEPX) and asthma symptoms, lung function, and other markers of eosinophilic airway inflammation in asthmatic school children. *Methods.* A cross-sectional study was performed in 180 steroid dependent atopic children with stable moderately severe asthma, who were stable on 200 or 500 µg of fluticasone per day. uEPX was measured in a single sample of urine and was normalized for creatinine concentration (uEPX/c). Symptom scores were kept on a diary card. FEV<sub>1</sub> and PD<sub>20</sub> methacholine were measured. Sputum induction was performed in 49 and FE<sub>NO</sub> levels measured in 24 children. *Results.* We found an inverse correlation between uEPX/c and FEV<sub>1</sub> ( $r = -.20$ ,  $P = .01$ ) and a borderline significant correlation between uEPX/c and PD<sub>20</sub> methacholine ( $r = -.15$ ,  $P = .06$ ). Symptom score, %eosinophils and ECP in induced sputum and FE<sub>NO</sub> levels did not correlate with uEPX/c. *Conclusion.* uEPX/c levels did not correlate with established markers of asthma severity and eosinophilic airway inflammation in atopic asthmatic children.

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## 1. INTRODUCTION

Eosinophilic airway inflammation is the pathological substrate of allergic asthma both in adults and in children [1, 2]. The severity of airway inflammation correlates poorly with symptoms and lung function [3]. As asthma treatment with inhaled steroids aims at reducing inflammation, there is a need to monitor the disease with a marker of inflammation [4, 5]. Potential markers are serum eosinophilic cationic protein (ECP), induced sputum cellularity and soluble markers [6], and the concentration of nitric oxide in exhaled air (FE<sub>NO</sub>) [7, 8].

Eosinophil protein X (EPX) is one of the toxic proteins present in eosinophil granules and is released by activated eosinophils. EPX can be measured accurately in urine (uEPX) [9]. Therefore, uEPX can be regarded as a marker of eosinophil degranulation in vivo [10]. uEPX levels in allergic asthmatic children were found to be significantly higher than in healthy controls [11–14]. Treatment with inhaled steroids reduced uEPX levels [14]. We hypothesized that measuring

EPX in urine could potentially prove to be useful for monitoring eosinophilic airway inflammation in children and may complement other markers of asthma control such as symptom scores and lung function

The aim of this study was to evaluate the relationship between uEPX and current symptoms and lung function parameters, and the relation between uEPX, induced sputum eosinophilia, and FE<sub>NO</sub>. For this purpose, we analyzed cross-sectional data obtained at enrolment for a multicentre trial.

## 2. METHODS

### 2.1. Subjects

Data were obtained from steroid-dependent asthmatic children who took part in a large randomized controlled multicentre trial (CATO: Children Asthma Therapy Optimal). One hundred and eighty atopic (RAST  $\geq$  class 1 for at least one airborne allergen) children, median age 10.3 years (range 6–16 years), with a documented clinical history of

moderately severe asthma were recruited from paediatric clinics in 8 general hospitals and 7 university hospitals in The Netherlands. All had been treated with inhaled corticosteroids (ICS) for at least 4 weeks. Data were obtained during a clinic visit at the end of the run in period of 4–12 weeks. During this period, they were treated with fluticasone dipropionate 200  $\mu\text{g}/\text{d}$  ( $n = 102$ ) or 500  $\mu\text{g}/\text{d}$  ( $n = 78$ ). All parents and children if  $> 12$  years gave their written informed consent. The study was approved by the medical ethics committees of all participating hospitals.

## 2.2. Symptom scores

Two weeks before visiting the hospital, patients kept a diary in which symptoms (shortness of breath, wheeze, and cough) were scored twice a day each on a 4-point (0–3) scale. Cumulative symptom scores were calculated over 14 days (maximum score 252).

## 2.3. Fractional exhaled nitric oxide

The fractional concentration of exhaled nitric oxide ( $\text{FE}_{\text{NO}}$ ) was measured with the online single breath method, using the NIOX NO-analyzer (Aerocrine, Stockholm, Sweden) according to ERS/ATS guidelines [15].

As  $\text{FE}_{\text{NO}}$  could only be measured in 1 participating university centre, only part of the children underwent  $\text{FE}_{\text{NO}}$  measurements.

## 2.4. Flow-volume curves

Flow-volume curves and forced expiratory volume in 1 second ( $\text{FEV}_1$ ) were measured on a dry rolling seal spirometer according to recommendations [16]. Results are expressed as percentage of predicted values [17].

## 2.5. Bronchial challenge test

Bronchial responsiveness was determined by a methacholine challenge [18].  $\text{PD}_{20}$  methacholine (provocative dose of methacholine causing  $\text{FEV}_1$  fall 20% from baseline) was assessed by linear interpolation of the last two points of the log dose-response curve where  $\text{FEV}_1$  had fallen below 20% of baseline value.

## 2.6. Sputum induction and processing

Sputum induction was performed by 5 university centres and 3 paediatric clinics in general hospitals. Sputum was induced according to a standardized method by inhaling an aerosol prepared from hypertonic sodium chloride 4.5% w/v [19, 20]. Differential cell counts of the cytopspins were performed by counting 500 cells. Sputum samples containing more than 80% squamous cells were excluded from the analysis [20].

In sputum supernatant, ECP was measured by fluoroenzyme immunoassay (Pharmacia, Uppsala, Sweden)

## 2.7. Urinary eosinophil protein X

A spot sample urine was collected from each individual at the clinic visit and immediately stored at  $-20^\circ\text{C}$ . uEPX was determined using a commercial enzyme-linked immunosorbent assay (ELISA) for human EPX in 50-fold diluted samples according to the manufacturers recommendations (Medical and Biological Laboratories, Naka-Ku Nagoya, Japan). The sensitivity of the assay was 0.62 ng/mL. Urinary creatinine levels were measured by using the alkaline picrate method (Jaffé reaction) (Roche, Mannheim, Germany). Urinary EPX concentrations were expressed as  $\mu\text{g}$  per mmol creatinine (uEPX/c).

## 2.8. Data analysis

All variables with a non-Gaussian distribution (symptom score,  $\text{PD}_{20}$  methacholine,  $\text{FE}_{\text{NO}}$ , % eosinophils in sputum, ECP in sputum, and uEPX) could be normalized by log-transformation. The significance of the relation between uEPX and lung function variables or other markers of inflammation was calculated using Spearman's rank correlation coefficients. A two-sided  $P$  value of  $<.05$  was considered statistically significant.

## 3. RESULTS

One hundred and eighty subjects (105 boys (58.3%)) participated. Asthma was controlled by fluticasone dipropionate 200  $\mu\text{g}/\text{day}$  ( $n = 102$ ) or 500  $\mu\text{g}/\text{day}$  ( $n = 78$ ).

All subjects performed spirometry and recorded symptoms in a diary. Six children inhaled short-acting  $\beta$ -agonists prior to the visit, their results were excluded from analysis. One hundred and seventy eight children performed a bronchial challenge test; two had  $\text{FEV}_1 < 80\%$  of personal best and were therefore not tested. Children who had used  $\beta$ -agonist within 8 hours before the test ( $n = 6$ ) were again excluded. For logistic reasons, sputum induction was done in part of the subjects. Forty nine of the 98 sputum inductions yielded adequate sputum samples (50%). At randomization, only one university centre had the facility to measure  $\text{FE}_{\text{NO}}$  ( $n = 24$  subjects).

Baseline results of lung function, symptom score, and markers of inflammation are given in Table 1. uEPX/c showed a log-normal distribution, median 185  $\mu\text{g}/\text{mmol}$  creatinine (range 2–3114  $\mu\text{g}/\text{mmol}$  creatinine). UEPX/c did not correlate with age and was not different between boys and girls.

### 3.1. Relation between uEPX/c and clinical markers of asthma severity (Table 2)

UEPX/c did not correlate with symptom scores or inhaled steroid dose. There was a significant inverse correlation of uEPX/c with  $\text{FEV}_1$  ( $r = -.18$ ,  $P = .02$ ) (Figure 1). The association between uEPX/c and  $\text{FEV}_1$  did not significantly differ between children using 200  $\mu\text{g}$  fluticasone per day and those using 500  $\mu\text{g}$  (Anova,  $P = .19$ ). For each 10% points increase

TABLE 1: Characteristics of study subjects. Values are median (range). FEV<sub>1</sub> is forced expiratory volume in 1 second; PD<sub>20</sub> methacholine is provocative dose of methacholine causing FEV<sub>1</sub> fall 20% from baseline; ECP is eosinophil cationic protein; FE<sub>NO</sub> is fractional concentration of nitric oxide in exhaled air; uEPX/c is urinary eosinophil protein X per mmol creatinine.

	Fluticasone dose		Total
	200 µg/day	500 µg/day	–
Age (years)	10(96.4 – 16.8) <i>n</i> = 102	11.3(6.4 – 16.7) <i>n</i> = 78	10.3(6.4 – 16.8) <i>n</i> = 180
Gender (m/f)	60/40	45/33	105/75
FEV <sub>1</sub> (pred. %)	99(56 – 135) <i>n</i> = 101	96(56 – 96) <i>n</i> = 73	97(56 – 135) <i>n</i> = 174
Cumulative symptom score	18.5(0 – 113) <i>n</i> = 102	14(0 – 152) <i>n</i> = 78	17.0(0 – 152) <i>n</i> = 180
PD <sub>20</sub> methacholine (µg)	200(3 – > 1570) <i>n</i> = 100	48(1 – > 1570) <i>n</i> = 72	68(1 – > 1570) <i>n</i> = 172
Eosinophils sputum (%)	1(0 – 72) <i>n</i> = 29	1(0 – 43) <i>n</i> = 20	1.0(0 – 72) <i>n</i> = 49
ECP sputum (ng/ml)	17(0 – 2345) <i>n</i> = 24	38(0 – 538) <i>n</i> = 19	29(0 – 2345) <i>n</i> = 43
FE <sub>NO</sub> (ppb)	11(5 – 63) <i>n</i> = 12	9(1 – 29) <i>n</i> = 12	10(1 – 63) <i>n</i> = 24
uEPX/c (µg/mmol)	189(2 – 2828) <i>n</i> = 102	180(10 – 3114) <i>n</i> = 78	185(2 – 3114) <i>n</i> = 180

TABLE 2: Correlations between uEPX or uEPX-c and clinical markers of asthma severity or markers of asthmatic inflammation. *r* values were all analyzed by Spearman's rank correlation tests. uEPX/c is urinary eosinophil protein X per mmol creatinine; FEV<sub>1</sub> is forced expiratory volume in 1 second; PD<sub>20</sub> methacholine is provocative dose of methacholine causing FEV<sub>1</sub> fall 20% from baseline; ECP is eosinophil cationic protein; FE<sub>NO</sub> is fractional concentration of nitric oxide in exhaled air.

Variable	<i>N</i>	Log uEPX/c	
		<i>r</i>	<i>P</i>
Age	180	–.01	.90
Symptom score	180	.03	.72
FEV <sub>1</sub>	174	–.18	.02
PD <sub>20</sub> methacholine	172	–.14	.08
% eosinophils in sputum	49	.17	.26
ECP sputum	43	–.03	.83
FE <sub>NO</sub>	24	.16	.46

of FEV<sub>1</sub> (pred. %) the geometric mean EPX/c ratio decreases 18% (95% CI: 5,30%). The correlation between uEPX/c and PD<sub>20</sub> methacholine was borderline significant (*r* = –.14, *P* = .08).

### 3.2. Relation between uEPX and markers of asthmatic airway inflammation (Table 2)

uEPX/c did not correlate with the % eosinophils or ECP in induced sputum, or with FE<sub>NO</sub>. Relations between uEPX and PD<sub>20</sub> methacholine or markers of asthmatic airway inflammation did not significantly differ when analysis was adjusted for fluticasone dose.

Correlations were similar when children with eczema were excluded from the analysis.

## 4. DISCUSSION

We found a significant correlation of uEPX/c and FEV<sub>1</sub>, and no association between uEPX/c and bronchial responsiveness or symptom scores in a large group of children with moderately severe allergic asthma. In subgroups, no significant correlations between uEPX/c and other markers of eosinophilic airways inflammation (% eosinophils and ECP in induced sputum or FE<sub>NO</sub>) were found.

This is the first study reporting uEPX/c levels in relation with markers of asthma severity and inflammation in a large population of children with atopic asthma, treated with inhaled steroids. Lugosi et al. have shown that uEPX levels were increased in symptomatic versus nonsymptomatic children with asthma, treated with inhaled steroids or disodium

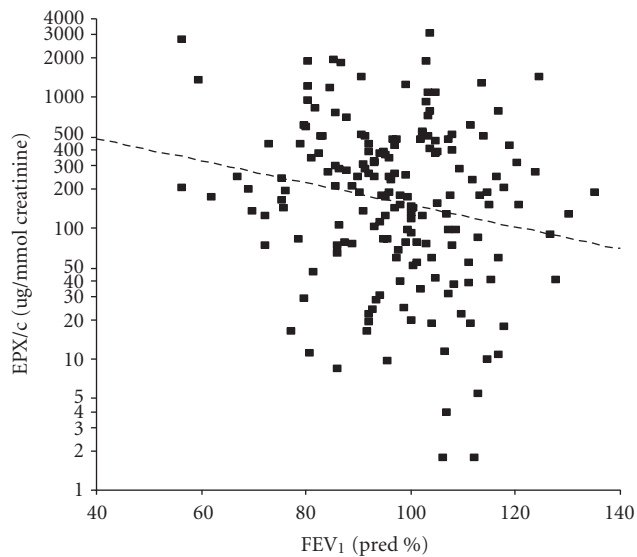


FIGURE 1: Scatter plot of urinary eosinophil protein X per mmol creatinine (uEPX-c) versus forced expiratory volume in 1 second ( $FEV_1$ ),  $n = 174$ .

cromoglycate [21], and Oosaki et al. [22] found significantly elevated uEPX levels during acute asthma exacerbations in children. All subjects included in our study had stable well-controlled asthma, as evidenced by a median cumulative symptom score of only 17 of a maximum of 252. Conflicting data have been published on the association between uEPX/c and pulmonary function tests [21, 23]. We found a significant negative correlation between  $FEV_1$  and uEPX/c. It should be mentioned that the scatter was wide, and individual uEPX/c therefore varied widely for a given  $FEV_1$  level. Hence, such correlations are unlikely to be detected in smaller groups. However, the within-subject variation of both parameters in time had not been studied. We confirmed our hypothesis that uEPX/c and bronchial hyperresponsiveness are not closely correlated. Lack of correlation between the severity of bronchial hyperresponsiveness and uEPX/c levels was also reported in 3 previous studies [24–26]. A close correlation between bronchial hyperresponsiveness and uEPX/c was not expected, because bronchial hyperresponsiveness is multifactorial and is not only caused by (eosinophilic) airways inflammation, but also by airway geometry, airway remodelling, and autonomic dysregulation.

Our hypothesis that uEPX/c would correlate with markers of eosinophilic airway inflammation could not be confirmed, as we found no correlation between uEPX/c and the percentage eosinophils in induced sputum. Others likewise found no correlation between uEPX/c and bronchoalveolar lavage cell counts in adult asthmatic patients [10]. An alternative explanation for not finding significant correlations between percentages of sputum eosinophils and uEPX/c could be that uEPX is only released by activated eosinophils, whereas in sputum we counted activated as well as nonactivated eosinophils. Also, the number of children from whom

suitable sputum samples or  $FE_{NO}$  values were obtained was relatively small.

We found no correlation between uEPX/c and sputum ECP levels. In contrast, Mattes et al. [11] reported a positive correlation between uEPX/c and sputum ECP in 25 stable asthmatic children on inhaled corticosteroids. They found much higher ECP concentrations than we did (median 453 ng/mL, range 40–2600; and 29 ng/mL, 1–2345, resp.). The reason for this is not clear, but may be related to different sputum processing techniques.

One could argue that the lack of correlation between uEPX/c and percentage of sputum eosinophils, or ECP levels in sputum supernatant, could be due to the wide scatter of uEPX. However, all urine samples were immediately stored at  $-20^{\circ}\text{C}$  and uEPX and urinary creatinine levels measurements were performed in a central laboratory (Leiden University Medical Hospital) to reduce variability in the analysis. All EPX measurements were done in duplicate and the within-subject reproducibility of uEPX levels was good.

It has been reported that in atopic dermatitis, concentrations of eosinophil-specific mediators, including uEPX/c, are increased [27, 28]. However, we found that the presence or absence of atopic eczema did not influence the correlations between uEPX/c and the percentage eosinophils or ECP in induced sputum. We cannot exclude that heterogeneity of study groups with respect to other atopic disorders than asthma could have affected the correlation between uEPX/c and other markers of eosinophilic airways inflammation.

At the onset of our study, a circadian rhythm of uEPX/c had not been reported. Urine samples were not all obtained at the same time of the day. Since the start of our study, it became evident that a circadian rhythm of uEPX/c with lowest levels at 7 p.m. and highest at 7 a.m. in both asthmatic and healthy controls exists [23, 29–31]. Hence, diurnal variability may have introduced scatter of uEPX, thus weakening a possible correlation.

Two previous studies reported significant positive correlations between uEPX/c and  $FE_{NO}$  in corticosteroid-dependent childhood asthma [11, 29]. We found no significant correlation between uEPX/c and  $FE_{NO}$  in a small subgroup of the study population. For  $FE_{NO}$ , no important circadian variation was found, employing the same measurement technique that we have used [32], but conflicting results have also been published [27, 33]. A possible circadian rhythm might have affected  $FE_{NO}$  and weakened any cross-sectional relationship.

In conclusion, the present data show a weak inverse correlation between uEPX/c and  $FEV_1$ , and a borderline correlation between uEPX/c and  $PD_{20}$  methacholine. No significant correlation was found between uEPX/c and markers of eosinophilic airway inflammation including % eosinophils or ECP levels in induced sputum or  $FE_{NO}$ . The number of children performing  $FE_{NO}$  was small, therefore this correlation should be interpreted with caution. Our findings are not encouraging for uEPX/c as a complementary marker of airway inflammation in asthma. As to whether uEPX/c can be useful as a marker for monitoring asthma management in children is worth prospectively looking at.



## APPENDIX

This work has been prepared by the authors on behalf of the CATO Study Group.

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## REFERENCES

- [1] W. Busse, S. P. Banks-Schlegel, and G. L. Larsen, "Childhood-versus adult-onset asthma," *American Journal of Respiratory and Critical Care Medicine*, vol. 151, no. 5, pp. 1635–1639, 1995.
- [2] Y. Cai, K. Carty, R. L. Henry, and P. G. Gibson, "Persistence of sputum eosinophilia in children with controlled asthma when compared with healthy children," *The European Respiratory Journal*, vol. 11, no. 4, pp. 848–853, 1998.
- [3] A. Jatakanon, S. Lim, S. A. Kharitonov, K. F. Chung, and P. J. Barnes, "Correlation between exhaled nitric oxide, sputum eosinophils, and methacholine responsiveness in patients with mild asthma," *Thorax*, vol. 53, no. 2, pp. 91–95, 1998.
- [4] J. K. Sont, L. N. A. Willems, E. H. Bel, et al., "Clinical control and histopathologic outcome of asthma when using airway hyperresponsiveness as an additional guide to long-term treatment," *American Journal of Respiratory and Critical Care Medicine*, vol. 159, no. 4, part 1, pp. 1043–1051, 1999.
- [5] R. H. Green, C. E. Brightling, S. McKenna, et al., "Asthma exacerbations and sputum eosinophil counts: a randomised controlled trial," *The Lancet*, vol. 360, no. 9347, pp. 1715–1721, 2002.
- [6] P. G. Gibson, "Use of induced sputum to examine airway inflammation in childhood asthma," *Journal of Allergy and Clinical Immunology*, vol. 102, no. 5, pp. S100–S101, 1998.
- [7] G. L. Piacentini, A. Bodini, S. Costella, et al., "Exhaled nitric oxide and sputum eosinophil markers of inflammation in asthmatic children," *The European Respiratory Journal*, vol. 13, no. 6, pp. 1386–1390, 1999.
- [8] P. G. Gibson, R. L. Henry, and P. Thomas, "Noninvasive assessment of airway inflammation in children: induced sputum, exhaled nitric oxide, and breath condensate," *The European Respiratory Journal*, vol. 16, no. 5, pp. 1008–1015, 2000.
- [9] C. M. Reimert, U. Minuva, A. Kharazmi, and K. Bendtzen, "Eosinophil protein X/eosinophil derived neurotoxin (EPX/EDN). Detection by enzyme-linked immunosorbent assay on a purification from normal human urine," *Journal of Immunological Methods*, vol. 141, no. 1, pp. 97–104, 1991.
- [10] V. Cottin, P. Deviller, F. Tardy, and J.-F. Cordier, "Urinary eosinophil-derived neurotoxin/protein X: a simple method for assessing eosinophil degranulation in vivo," *Journal of Allergy and Clinical Immunology*, vol. 101, no. 1, part 1, pp. 116–123, 1998.
- [11] J. Mattes, K. Storm van's Gravesande, U. Reining, et al., "NO in exhaled air is correlated with markers of eosinophilic airway inflammation in corticosteroid-dependent childhood asthma," *The European Respiratory Journal*, vol. 13, no. 6, pp. 1391–1395, 1999.
- [12] C. Severien, A. Artlich, S. Jonas, and G. Becher, "Urinary excretion of leukotriene E<sub>4</sub> and eosinophil protein X in children with atopic asthma," *The European Respiratory Journal*, vol. 16, no. 4, pp. 588–592, 2000.
- [13] M. O. Hoekstra, H. Hovenga, J. Gerritsen, and H. F. Kauffman, "Eosinophils and eosinophil-derived proteins in children with moderate asthma," *The European Respiratory Journal*, vol. 9, no. 11, pp. 2231–2235, 1996.
- [14] S. Kristjánsson, I.-L. Strannegård, Ö. Strannegård, C. Petersson, I. Enander, and G. Wennergren, "Urinary eosinophil protein X in children with atopic asthma: a useful marker of anti-inflammatory treatment," *Journal of Allergy and Clinical Immunology*, vol. 97, no. 6, pp. 1179–1187, 1996.
- [15] E. Baraldi, J. C. de Jongste, B. Gaston, et al., "Measurement of exhaled nitric oxide in children, 2001," *The European Respiratory Journal*, vol. 20, no. 1, pp. 223–237, 2002.
- [16] P. H. Quanjer, G. J. Tammeling, J. E. Cotes, O. F. Pedersen, R. Peslin, and J. C. Yernault, "Lung volumes and forced ventilatory flows. Report Working Party Standardization of lung function tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society," *The European Respiratory Journal, Supplement*, vol. 16, pp. 5–40, 1993, see comments.
- [17] A. Zapletal, M. Samanek, and T. Paul, "Lung function in children and adolescents. Methods and reference values," in *Progress in Respiration Research*, vol. 22, pp. 114–218, Karger, Basel, Switzerland, 1987.
- [18] D. Birnie, G. W. S. Thoe Schwartzberg, W. C. J. Hop, E. E. M. van Essen-Zandvliet, and K. F. Kerrebijn, "Does the outcome of the tidal breathing and dosimeter methods of assessing bronchial responsiveness in children with asthma depend on age?" *Thorax*, vol. 45, no. 3, pp. 199–202, 1990.

- [19] A. Efthimiadis, A. Spanevello, Q. Hamid, et al., "Methods of sputum processing for cell counts, immunocytochemistry and in situ hybridisation," *The European Respiratory Journal, Supplement*, vol. 20, no. 37, pp. 19s–23s, 2002.
- [20] J. C. C. M. In't Veen, H. W. F. M. de Gouw, H. H. Smits, et al., "Repeatability of cellular and soluble markers of inflammation in induced sputum from patients with asthma," *The European Respiratory Journal*, vol. 9, no. 12, pp. 2441–2447, 1996.
- [21] E. Lugosi, G. Halmerbauer, T. Frischer, and D. Y. Koller, "Urinary eosinophil protein X in relation to disease activity in childhood asthma," *Allergy*, vol. 52, no. 5, pp. 584–588, 1997.
- [22] R. Oosaki, Y. Mizushima, A. Kawasaki, H. Mita, K. Akiyama, and M. Kobayashi, "Correlation among urinary eosinophil protein X, leukotriene E<sub>4</sub>, and 11-dehydrothromboxane B<sub>2</sub> in patients with spontaneous asthmatic attack," *Clinical and Experimental Allergy*, vol. 28, no. 9, pp. 1138–1144, 1998.
- [23] A. Labbé, B. Aublet-Cuvelier, L. Jouaville, et al., "Prospective longitudinal study of urinary eosinophil protein X in children with asthma and chronic cough," *Pediatric Pulmonology*, vol. 31, no. 5, pp. 354–362, 2001.
- [24] G. L. Piacentini, C. Peterson, D. G. Peroni, A. Bodini, and A. L. Boner, "Allergen avoidance at high altitude and urinary eosinophil protein X," *Journal of Allergy and Clinical Immunology*, vol. 104, no. 1, pp. 243–244, 1999.
- [25] M. O. Hoekstra, M. H. Grol, K. Bouman, et al., "Fluticasone propionate in children with moderate asthma," *American Journal of Respiratory and Critical Care Medicine*, vol. 154, no. 4, part 1, pp. 1039–1044, 1996.
- [26] D. Y. Koller, G. Halmerbauer, T. Frischer, and B. Roithner, "Assessment of eosinophil granule proteins in various body fluids: is there a relation to clinical variables in childhood asthma?" *Clinical and Experimental Allergy*, vol. 29, no. 6, pp. 786–793, 1999.
- [27] K. Oymar and R. Bjerknes, "Urinary eosinophil protein X in children with atopic dermatitis: relation to atopy and disease activity," *Allergy*, vol. 55, no. 10, pp. 964–968, 2000.
- [28] N. Pucci, E. Lombardi, E. Novembre, et al., "Urinary eosinophil protein X and serum eosinophil cationic protein in infants and young children with atopic dermatitis: correlation with disease activity," *Journal of Allergy and Clinical Immunology*, vol. 105, no. 2, pp. 353–357, 2000.
- [29] J. Mattes, K. Storm van's Gravesande, C. Moeller, M. Moseler, M. Brandis, and J. Kuehr, "Circadian variation of exhaled nitric oxide and urinary eosinophil protein X in asthmatic and healthy children," *Pediatric Research*, vol. 51, no. 2, pp. 190–194, 2002.
- [30] K. Storm van's Gravesande, J. Mattes, T. Grüntjens, et al., "Circadian variation of urinary eosinophil protein X in asthmatic and healthy children," *Clinical and Experimental Allergy*, vol. 29, no. 11, pp. 1497–1501, 1999.
- [31] M. Schosser, P. Hoeppe, K. Radon, and D. Nowak, "Circadian rhythm of U-EPX," *Allergy*, vol. 56, no. 8, pp. 792–793, 2001.
- [32] S. A. Kharitonov, F. Gonio, C. Kelly, S. Meah, and P. J. Barnes, "Reproducibility of exhaled nitric oxide measurements in healthy and asthmatic adults and children," *The European Respiratory Journal*, vol. 21, no. 3, pp. 433–438, 2003.
- [33] M. W.H. Pijnenburg, S. E. Floor, W. C.J. Hop, and J. C. De Jongste, "Daily ambulatory exhaled nitric oxide measurements in asthma," *Pediatric Allergy and Immunology*, vol. 17, no. 3, pp. 189–193, 2006.

